

Phytochemistry, GC-MS Analysis, physicochemical and *In vitro* Antimicrobial activity of seed and peel of *Citrus sinensis*

Smita Singh^{1*} and Nabya Nehal

Department of Biotechnology, Faculty of Engineering & Technology, Rama University,
Kanpur-209217 (India)

Email : smitas090@gmail.com; Mobile: +91-8957142914

^{1*}Corresponding Author

Abstract

Citrus fruits are one of the important fruit trees grown worldwide and appreciated for their refreshing juice and health benefits. The research paper has in depth literature review on the importance of *Citrus* fruits and investigate the numerous therapeutic properties of *Citrus* fruits. The research paper summarizes the findings of the phenolic content, Antioxidant activity of extracts of *Citrus* fruits on 10 samples. The research made to estimate the mineral composition (g 100-1), proximate composition (%) and food energy (g Cal-1), phytonutrients content of *Citrus* species on dry weight basis expressed as mg/100g , Vitamin composition of *Citrus* juice expressed as mg/100g for 5 species and tabulated the data with graphical representation. Finally, paper concluded the importance of *Citrus* fruits on the basis of results of antioxidant activity, total phenolic and total flavonoid contents of *Citrus* leaves. The research also find out the different parameter by vitro antimicrobial activity the oil samples of seeds and peels of orange and analysis the findings.

Citrus fruits are the main fruit trees grown throughout the world and are well – appreciated for their refreshing juice and health benefits. Numerous therapeutic properties have been attributed to *Citrus* fruits, like anticancer, antiviral, anti-tumor, anti-inflammatory activities, and effects on capillary fragility as well as an ability to inhibit platelet aggregation. Though there were many studies on antioxidant and antibacterial effect of juice and

edible parts, there are meager literature on the wastes of *Citrus* fruits of lemon and oranges of different varieties.

The clinical use of lemon and orange are investigated by identification of multiple constituents Alkaloids, polysaccharides, lectins, glycosides, phenolic compounds, flavonoids, anthocyanins, tannins, and sterols for antioxidants properties and d-limonene for better pharma-

cokinetic and pharmacodynamics properties^{3,29}. The *Citrus* peel wastes are rich in nutrients and contain many phytochemicals. These wastes can be effectively disposed by manufacturing by-product from them. The hydro-distillation and GC-MS analysis on by-product of orange peels made to find the aromatic compounds in the peel oil. The parameters of oil like limonene (86.75%), linalool (1.96%), α -pinene (1.63%), trans-limonene oxide (1.39%), and γ -terpinene (1.03%) confirm its utilization as food preservative^{1,6,7,8}. The food, cosmetic and chemical industries use The Cold pressed and microwave roasting of orange (*Citrus sinensis*) seeds oils studied and found the availability of fatty acids : linoleic, palmitic and oleic acids, phenolics, α -tocopherol, β -sitosterol conclude the application of oil in the food, cosmetic or chemical industries^{9,14,15,29}.

The experiment of CSE, MAE and UAE and SC-CO₂ extraction claim that antioxidant activity (DPPH method) and its correlation to TPC, TFC or individual flavonoids. Further the findings of total carotenoids, total carotenoids (19.01 mg/kg), total phenolic compounds (4.43 g/kg), α -tocopherol (135.65 mg/kg) and phytosterols (1304.2 mg/kg) form Extracted from orange (*Citrus sinensis*) seeds determine their antioxidant activity^{20,23}. Phytochemicals and assess antioxidant capacity from *Citrus* processing by-product and essential oils were tested *in vitro* for their anticandidial activity^{10,19}. Further Flavonoid-rich chloroform fraction (FRCF), microwave assisted extraction, MAE and supercritical CO₂ extraction, SC-CO₂) on the total phenol contents, total flavonoid contents, individual flavonoids, vitamin C and antioxidant activity

of orange peel confirm the antioxidant activity^{24-26,28}.

The present study has shown the usefulness of the extraction methodologies adopted for efficient extraction, processing and utilization of these *Citrus* fruit peel wastes and to characterize the phytochemicals, antioxidant property and antibacterial activities of fruit peel wastes of lemon and oranges.

Collection and identification of Citrus fruit peels :

Peels of *Citrus* fruits was collected from local *Citrus* juice shop. Further the *Citrus* Seed oil extracted by hydro distillation (3-4 hours extraction) and kept in fridge. The experimental process to find Physico-chemical parameters mentioned in Table-1.

Collection and preparation of sample seed & peel oil:

Orange seed oil : Seed manually separated and dry in air, grinded on electrical grinder machine approx. 1.5 kg of *Citrus sinensis* seed kept in flask. The n-hexane added as extracting solvent and find oil by rotary evaporated method.

Orange peel oil : Peels washed with double distilled water and dried in an oven at (105°C \pm 2) for two days to constant weight. The dried peels were ashed in a porcelain container placed in furnace for 6 h by stepwise increase of the temperature up to 500°C. The ashed samples were homogenized in porcelain mortar and grinder and filtered. Then same method as above for oil extraction.

Table-1. Experimental process to find Physico-chemical parameters of the oil samples

SNo.	Investigation Method	Parameters and Process
1	Physicochemical and Phytochemical Investigation of <i>Citrus</i> EO	Including refractive index, optical rotation, specific gravity, color, odor and solubility.
2	GC-MS Analysis of <i>Citrus</i> EO	Screening of bioactive compounds
3	DPPH Assay Antioxidant potential methanolic and shaken well.	500 μ L essential oil added in 3 ml of 0.002%

Table-2. *In vitro* Antimicrobial Activity the oil samples of seeds and peels of orange

SNo.	Investigation	Parameter to Checked
1	Physical properties of the oils	Refractive index ,specific gravity, flash and smoke points, cloud and pour points and viscosity test
2	Chemical properties of the oils	acid value (AV), free fatty acid (FFA), Iodine value (IV), saponification value (SV) and peroxide value (PV)
3	Determination of fatty acid composition	Fatty Acid composition
4	Total fatty matter (TFM)	determined by the petroleum spirit extraction method
5	Determination of total alkali	Total alkali obtained by method of AOAC (1990).
6	Foamability test	Measure height of the foam in the solution

Statistical analysis :

reported as mean values \pm SD with 95% confidence level. All experimental value are

The oil samples analyses were performed for three times, and the results were

summarized in different table heading from Table-3 to Table-10.

Table-3. Physico-chemical parameters of the oil samples.

Variable	Orange seed oil	Orange peel oil
pH	4.5	5.5
Colour	Golden yellowish	Brownish-yellow
Percentage yield (%)	40	32
Specific density (g/cm^3)	0.95	0.75
Cloud point (degree C)	14	15
Pour point (degree C)	7.5	9
Viscosity 100°C (CST)	3.25	0.95
Refractive Index	1.4	1.48
Acid value (mgKOH/g)	23.5	26.1
Peroxide value (mgKOH/g)	18.2	5.5
Free fatty acid (% as oleic acid)	11.5	111.5
Saponification value (mgKOH/g)	221.5	42.5

The experimental values of Orange seed and peel oil are summarized in above tables addressed that excess of fatty acids make Seed oil is more acidic than the peel oil. Carotenoids responsible for yellow colour and higher saponification value make it suitable for to use in edible product. The observations like the smoke point, flash point and free fatty acid content of the oils have a linear relationship and peroxide value serves as a common indicator of lipid oxidation.

Table-4. Phytonutrients content of *Citrus* species on dry weight basis expressed as mg/100g.

Species	Alkaloids	Flavonoids	Tannins	Phenols	Saponins
<i>Blanco C. reticulata</i>	0.4±0.15	0.3±0.05	0.03±0.01	0.035±0.02	0.035±0.02
<i>C. aurantifolia</i>	0.35±0.15	0.3±0.05	0.05±0.01	0.025±0.01	0.25±0.1
<i>C. limonum</i>	0.6±0.25	0.6±0.10	0.02±0.01	0.055±0.01	0.5±0.05
<i>C. vitis</i>	0.65±0.15	0.25±0.1	0.03±0.15	0.09±0.01	0.25±0.05
<i>C. sinensis</i>	0.6±0.15	0.2±0.05	0.04±0.05	0.015±0.01	0.06±0.02

Table-5. Proximate composition (%) and food energy (g Cal⁻¹) values of *Citrus* species

Species	Moisture	Crude protein	Crude fiber	Ash
<i>C. reticulata</i>	5.6±0.15	11.9±0.02	5.35±0.15	4.5±0.15
<i>C. aurantifolia</i>	6.1±0.25	10.5±0.15	6.25±0.25	7.6±0.15
<i>C. limonum</i>	6.5±0.15	14.5±0.15	6.85±0.15	5.5±0.15
<i>C. vitis</i>	6.35±0.25	12.95±0.25	7.4±0.25	5.2±0.15
<i>C. sinensis</i>	4.5±1.15	18.15±0.25	5.66±0.2	5.52±0.25

Table-6. Mineral composition (g 100⁻¹) *Citrus* species

Species	P	K	Mg	Na	Ca
<i>C. reticulata</i>	0.3±0.15	0.48±0.22	0.5±0.12	0.35±0.25	2.5±0.25
<i>C. aurantifolia</i>	0.3±0.2	1.5±0.15	0.4±0.15	0.38±0.1	2.9±0.15
<i>C. limonum</i>	0.4±0.25	0.4±0.15	0.35±0.01	0.35±0.02	2.2±0.15
<i>C. vitis</i>	0.25±0.15	0.3±0.02	0.31±0.15	0.3±0.15	3.5±0.15
<i>C. sinensis</i>	0.45±0.25	0.85±0.1	0.35±0.1	0.34±0.25	2.12±0.15

Table-7. Vitamin composition of *Citrus* juice expressed as mg/100g

Species	Ascorbic Acid	Thiamine	Riboflavin	Niacin
	Vitamin C	Vitamin B ₁	Vitamin B ₁	Nicotinic acid
<i>C. reticulata</i>	32.5±0.25	0.15±0.1	0.02±0.1	0.45±0.11
<i>C. aurantifolia</i>	23.5±0.15	0.15±0.15	0.05±0.15	0.04±0.2
<i>C. limonum</i>	62.5±0.2	0.9±0.2	0.03±0.08	0.15±0.35
<i>C. vitis</i>	35.89±0.25	0.1±0.01	0.08±0.15	0.34±0.2
<i>C. sinensis</i>	20.5±0.2	0.06±0.1	0.15±0.15	0.35±0.1

Table-8. Percentage yield of *Citrus* essential oils by fabricated unit

Citrus species Essential oils (EO)	Raw material Input (g)	Time extract (minutes)	Oil volume (ml)	Productivity (ml/2000g)%
<i>C. paradisi</i>	2000	200	8.8	0.44
<i>C. sinensis var. Malta</i>	2000	220	7.5	0.375
<i>C. reticulata var. Mandarin</i>	2000	215	6.5	0.325
<i>C. sinensisavar Mousami</i>	2000	240	6.2	0.31
<i>C. reticulata var. Tangerine</i>	2000	225	5.8	0.29

Table-9. The phenolic content and Antioxidant activity of extracts of *Citrus* fruits

Sample	Total phenolic content mg GA/mL	IC ₅₀ (mg/mL)
Lemon peel	0.5±0.008	19.85±2
Lemon juice	0.35±0.02	78.15±5.5
Orange peel	0.48±0.025	20.2±0.25
Orange juice	0.45±0.015	6.5±0.5

Table-10. Total phenolic and total flavonoid contents of *Citrus* leaves

Extracts	Total phenolic content, mg, Gallic acid equivalents/g DM	Fls vonoids, mg Quercetin equivalents/g DM
Aqueous extract		
<i>C. cleamentina</i>	124.85±0.25	46.8±2.5
<i>C. aurantifolia</i>	105.95±2.5	38.2±1.5
<i>C. limon</i>	98.52±3.05	37.85±1.5
<i>C. navel</i>	85.68±8.1	20.5±2
<i>C. hamlin</i>	85.25±7.5	19.2±2.5
Methanolic extract		
<i>C. cleamentina</i>	11.5±0.5	8.2±0.5
<i>C. limon</i>	4.5±0.75	3±0.35
<i>C. navel</i>	7.5±0.65	4.1±0.4
<i>C. grandis</i>	3.1±0.08	1.05±0.2

Further the different Physicochemical value of Orange seed & peel Oil obtain from experiments are visualize in graphical representation from Figures-1 to Figure-6.

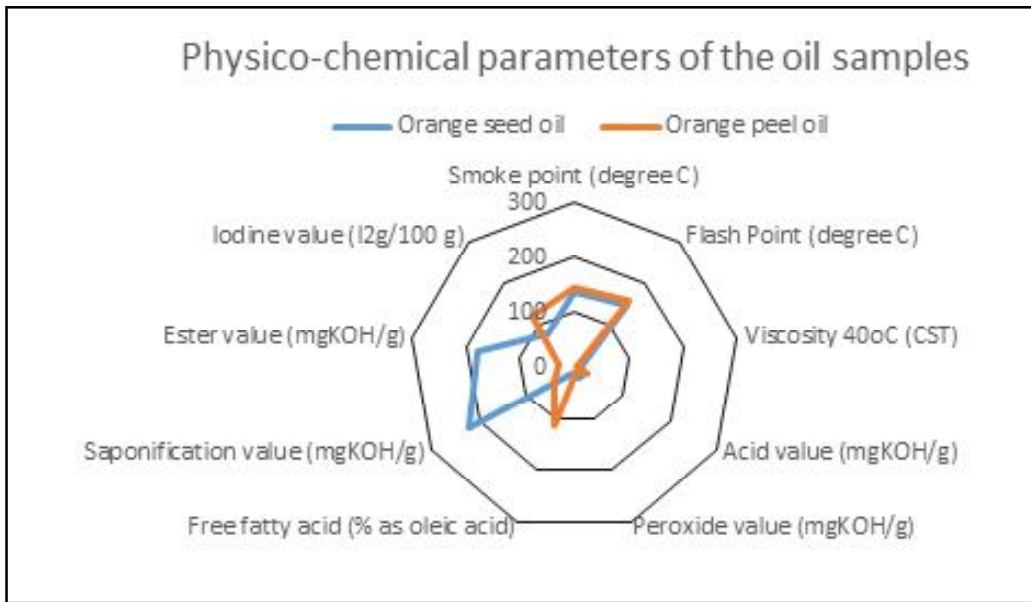


Figure-1. Graphical re-presentation of Physico-chemical parameters of the oil samples of orange seed and peel

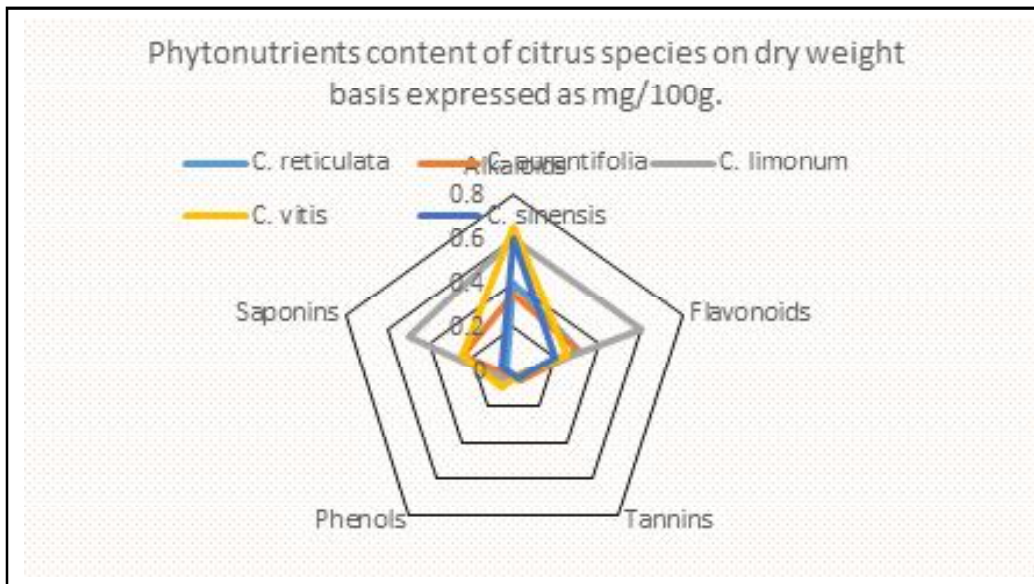


Figure-2: Graphical re-presentation of Phytonutrients content of citrus species

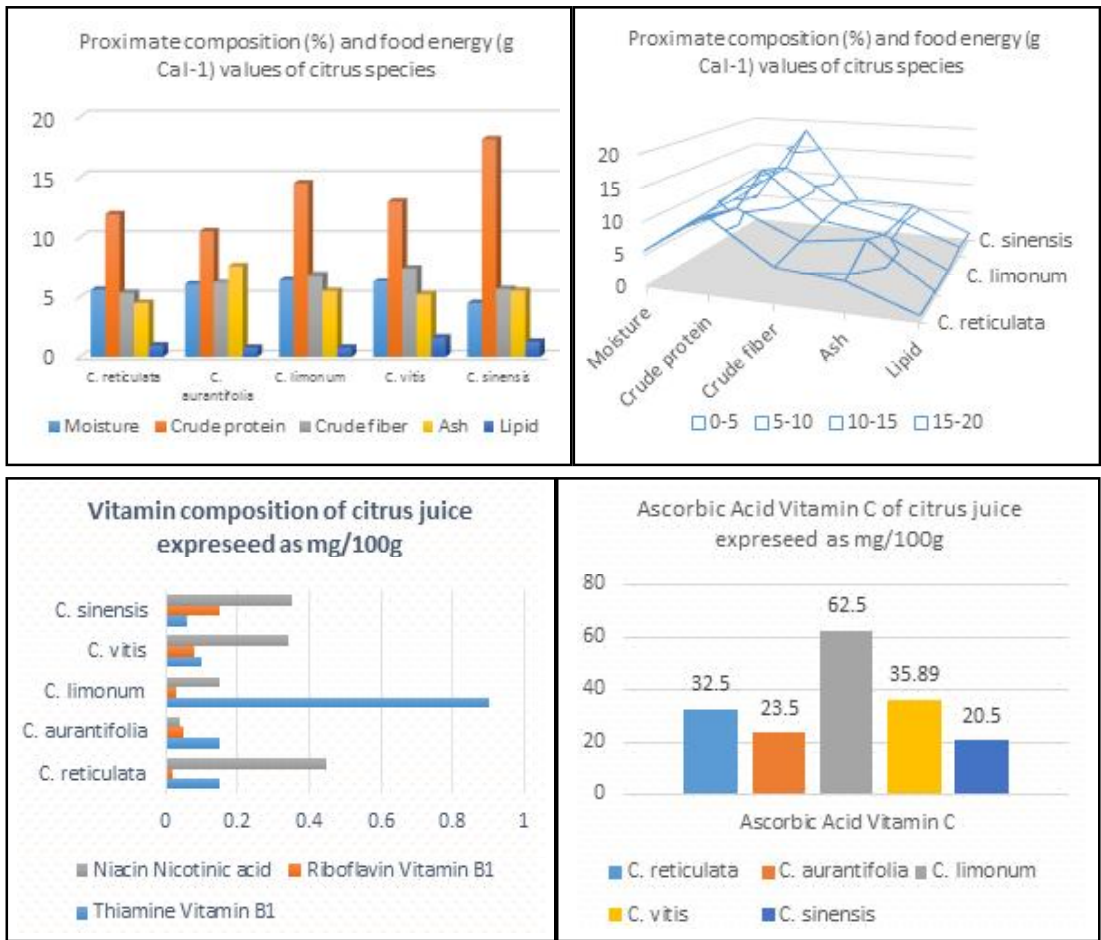


Figure-3. Graphical re-presentation of Chemical Composition of Citrus species

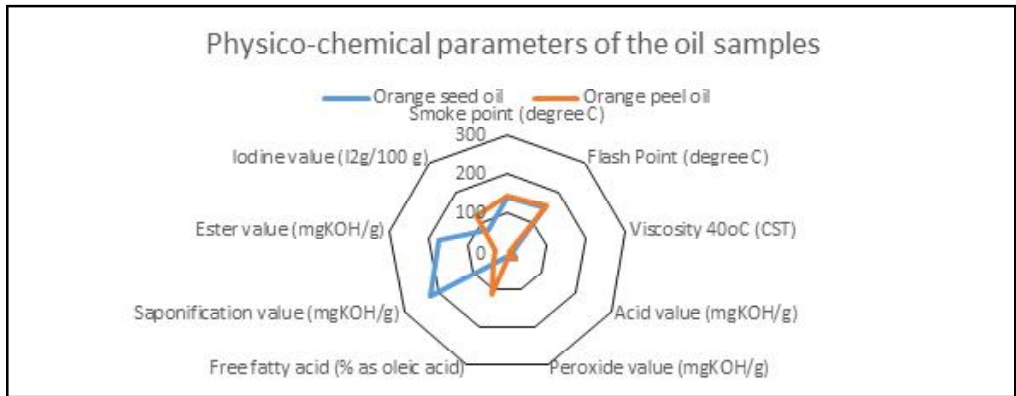


Figure-4. Physico-chemical parameters of the oil samples of Orange seed and peel oil

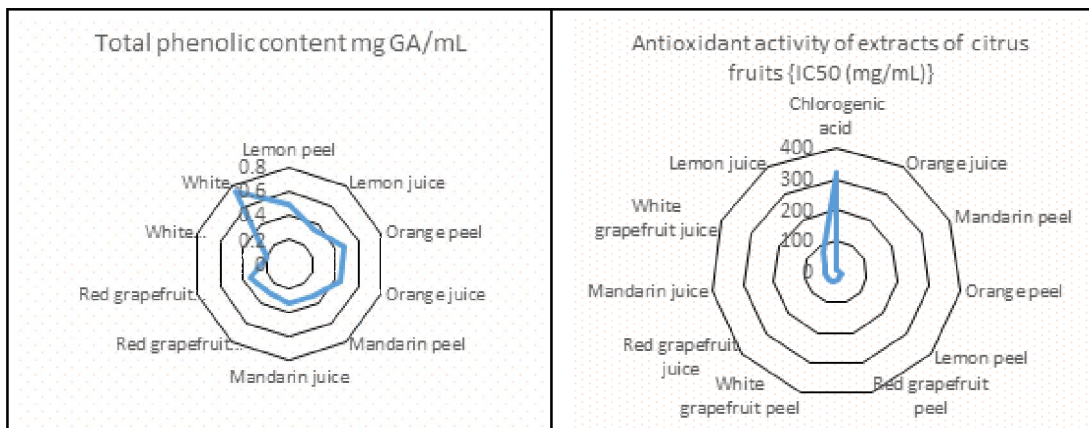


Figure-5. Total phenolic content and antioxidant activity of citrus fruits.

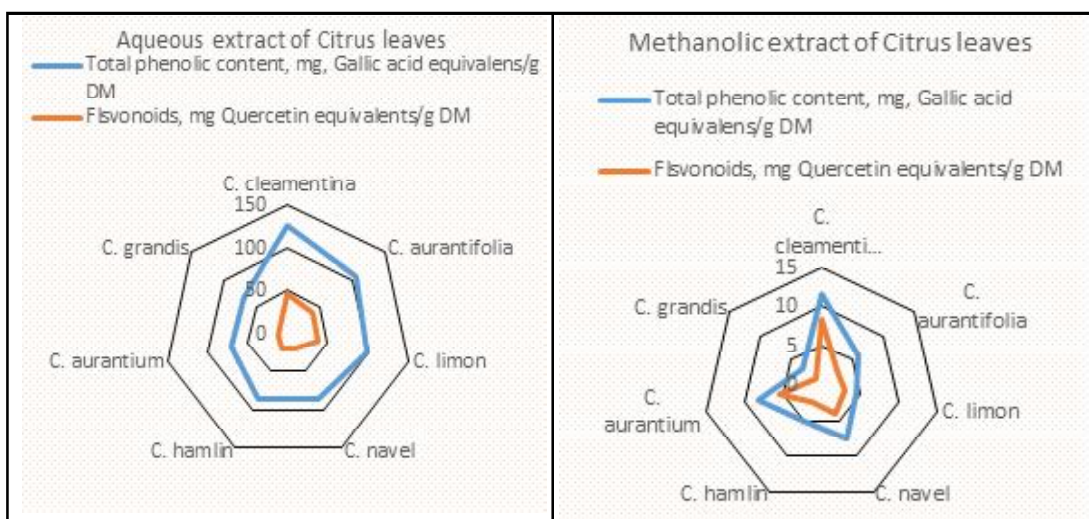


Figure-6. Aqueous and Methanolic Extract Citrus leaves property

The research on the seed and peel of *Citrus Sinensis* had made in different angle, the physicochemical and phytochemical investigation, GC-MS Analysis and DPPH Assay Antioxidant potential. The values of Physico-chemical parameters of the oil samples of Orange peel oil is higher than Orange seed oil (pH value 5.5 in respect of 4.5 , Acid value (mgKOH/g) is 26.1 in respect

of 23.5). Total phenolic and total flavonoid contents of Citrus leaves measure by aqueous extract and Methanolic extract along with other 6 parameter, further finding are summarized in tabulated and graphical representation for ease of understanding. The results of our study showed that *Citrus sinensis*, *C. limetta* and *C. limon* peel oil have the probability to be applied as a natural

constituent of food preservations, cosmetics and medi-cines as they exhibit a strong antioxidant, antibacterial and antifungal activity against food borne pathogens. In conclusion, the Chemical analysis of essential oil extracted from *Citrus sinensis*, *C. limetta* and *C. limon* showed in research able to justify the importance of the citrus seed oil.

The authors gratefully acknowledge the necessary Laboratory facilities and constant supervision provided by the Department of Biotechnology, Rama University, Kanpur, U.P., India for their generous support during the research work.

Conflict of interest :

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References :

1. Abdel- Fattah, Sh. M, H.A Yehia, A.S.M. Fouzy, Ramadan Manal M. and A. Nooh (2015) *International Journal of Advanced Research* 3(10): 1257–1269.
2. Abeyasinghe., D.C., L. Xian, S.C. De, Z.W. Shu, Z. C. Hua, and C. K. Song, (2007) *Journal of Food Chemistry* 104(4): 1338-1344. doi.org/10.1155/2017/8401074
3. Ahmed Abdullah Khan, Tarique Mahmood, Hefazat H. Siddiqui and Juber Akhtar (2016) *Journal of Chemical and Pharmaceutical Research* 8(3): 555-563.
4. Anwar, F., R. Naseer, M. I. Bhangar, S. Ashraf, F.N. Talpur, and F.A. Aladedunye (2008) *Journal of American Oil Chemist. Society* 85: 321-330. doi.org/10.1007/s11746-008-1204-3
5. Boluda, A. M. and G. A. Lopez, (2012) *Industrial Crops and Products* 41: 188-197 doi.org/10.1016/j.indcrop.2012.04.031
6. Bordoloi, A. K., M. G Pathak, J. Sperkova and P. A. Leclercq, (1999) *Journal of Essential Oil Research* 11(5): 629-632 doi.org/10.1080/10412905.1999.9701228
7. Breemen, R.B., L. Dong and N.D. Pajkovic (2012) *International Journal of Mass Spectrometry* 312: 163-172. doi.org/10.1016/j.ijms.2011.07.030
8. Cheong, M. W., Z. S. Chong, S. Q. Liu, W. Zhou, P. Curran, and B. Yu, (2012) *Food Chemistry* 134: 686–695 doi.org/10.1016/j.foodchem.2012.02.162
9. Combariza, M.Y., C.B. Tirado, E. Stashenko and T. Shibamoto, (1994) *Journal of High Resolution Chromatography* 17(9): 643-646 doi.org/10.1002/jhrc.1240170905
10. Dutta, B. K., S. Karmakar, A. Naglot, and M. Begam, (2007) *Journal of Mycoses* 50(2): 121-124 doi.org/10.1111/j.1439-0507.2006.01332.x
11. Furiga, A., F. A. Lonvaud, and C. Badet, (2009) *Food Chemistry* 11: 1037-140 doi.org/10.1016/j.foodchem.2008.08.059
12. Gualdani, R., M. M. Cavalluzzi, G. Lentini, and S. Habtemariam, (2016) *Molecules* 21(11): 1530 doi:10.3390/molecules21111530
13. Guneser, B. A., and E. Yilmaz, (2017a) *Journal of the American Oil Chemists' Society* 94(5): 723-731 DOI 10.1007/s11746-017-2977-z
14. Guneser, B. A., and E. Yilmaz, (2017) *Grasas y Aceites* 68(1): 175 doi: http://dx.doi.org/10.3989/gya.0800162
15. Güneşer, B. A., and E. Yilmaz, (2019) *Journal of food science and technology* 56(2): 634-642 doi.org/10.1007/s13197-018-3518-y

16. Gursoy, N., B. Tepe, and M. Sokmen, (2010) *International Journal of Food Properties* 13(5): 983-991 doi.org/10.1080/10942910902927136
17. Hamdan, D., M. Z. El-Readi, E. Nibret, F. Sporer, N. Farrag, A. El-Shazly, and M. Wink, (2010) *Pharmazie* 65(2): 141-148 doi.org/10.1691/ph2010.9731
18. Hamdan, D. I., R. H. Abdulla, M. E. Mohamed, and A. M. El-Shazly, (2013) *Journal of Pharmacognosy and Phytotherapy* 5(5): 83-90 DOI 10.5897/JPP2013.0277
19. Mehmood, T., M. R. Khan, M. A. Shabbir, and M. Anjum (2018) *Progress In Nutrition* 20: 279-288 DOI: 10.23751/pn.v20i1-S.6357
20. Mhiri N, I Ioannou, C Paris, M Ghoul and N Mihoubi Boudhrioua (2016) *Int J Food Nutr Sci.* 3(1): <https://doi.org/10.15436/2377-0619.16.789>
21. Nadia Mcharek and Belgacem Hanchi (2017) *Journal of Applied Botany and Food Quality* 90: 1 – 9 DOI: 10.5073/JABFQ.2017.090.001
22. Nasri, M., F. Bedjou, D. Porras, and S. Martínez-Flórez, (2017) *Phytothérapie*, 1-13 DOI 10.1007/s10298-017-1094-8
23. Neuza Jorge, Ana Carolina Da Silva And Caroline P. M. Aranha (2016) *Annals of the Brazilian Academy of Sciences An Acad Bras Cienc* 88(2): DOI: 10.1590/0001-3765201620140562
24. Nisha, S. N., A. A. Swedha, and J. S. N. Rahaman, (2013) *International Journal of Pharmaceutical Research and Bio-Science* 2(5): 1-13.
25. Nouha M'hiri, Ioannou Irina, Paris Cédric, Mohamed Ghoul and Nourhene Boudhrioua (2017) *Journal of Applied Pharmaceutical Science* 7 (11):126-135. DOI: 10.7324/JAPS.2017.71119
26. Priya E. Sanmuga, P. Senthamil Selvan and A. Syed Arshad (2018) *Current Science* 115(10):
27. Shabnam Javed, Ayesha Javaid, Shaista Nawaz, M. K. Saeed, Zaid Mahmood, S. Z. Siddiqui and Rauf Ahmad (2014) *Journal of Agricultural Science* 6(3) : DOI:10.5539/jas.v6n3p201
28. Sheng, W.Q., Y.N. Zou, and R.X. Xia (2006) *Euro. J. Soil. Bio.* 42: 166–172 doi: 10.3389/fmicb.2014.00682
29. Venkatalakshmi, P., V. Vadivel, and P. Brindha (2016) *International Journal of Green Pharmacy* 10 (1): DOI: <http://dx.doi.org/10.22377/ijgp.v10i1.600>
30. Yilmaz, E., and B. A. Güneşer, (2017) *Journal of food science and technology* 54(7): 1891-1900 DOI 10.1007/s13197-017-2622-8